ABSTRACT

Background: The contribution of indoor fungal exposure to childhood asthma is not completely clear.

Objective: To investigate airborne fungal flora within the homes of asthmatic and control children, and to assess the influence of housing characteristics regarding indoor fungi.

Methods: Forty-seven atopic asthmatic and 23 nonatopic control children were studied. Allergen sensitivity was determined by skin prick tests. A thorough assessment, using a questionnaire and inspection surveys, was carried out. Home visits were made between October 2000 and February 2001. Samples of airborne fungal spores were collected from four rooms by the “open Petri dish” method. Indoor temperature and humidity were measured.

Results: The total indoor fungal colony counts from the living rooms and bedrooms were significantly higher in the asthma group than in controls (p = .012 and p = .003, respectively). The most commonly isolated genus was Cladosporium. Twelve of the asthmatic patients (25.53 %) were found to be sensitive to fungal allergens. The factors found to be associated with indoor fungal growth in logistic regression were visible fungal patches in the bathrooms [(odds ratio (OR) = 5.75; 95 % CI 1.19 to 27.70)], and the age of the house [OR = 4.24; 95 % CI 1.34 to 13.45]. Total fungal colony numbers did not correlate with indoor temperature or humidity.

Conclusion: Fungal colony numbers were higher in the homes of asthmatic children than in those of controls. Therefore, indoor fungal exposure may contribute to childhood asthma. Bathrooms were the main source of fungal propagules. Old houses were more prone to fungal growth.


Indoor airborne fungal spores and home characteristics in asthmatic children from Edirne region of Turkey

M. Yazicioglu*, A. Asan*, U. Ones*, U. Vatansever*, B. Sen*, M. Ture*, M. Bostancioglu* and O. Pala*

*Associate Professor of Pediatrics. Department of Pediatrics. Trakya University Faculty of Medicine. Edirne. Turkey. †Professor in Biology. Department of Biology. Trakya University Faculty of Arts and Sciences. Edirne. Turkey. ‡Professor of Pediatrics. Allergy Clinic of the Department of Pediatrics. Istanbul University Faculty of Medicine. Istanbul, Turkey. §Assistant Professor of Pediatrics. Department of Pediatrics. Trakya University Faculty of Medicine. Edirne. Turkey. ¶Research Assistant in Biology. Department of Biology. Trakya University Faculty of Arts and Sciences. Edirne. Turkey. †Professor in Biostatistics. Department of Biostatistics. Trakya University. Edirne. Turkey. ‡Research Assistant in Pediatrics. Department of Pediatrics. Trakya University Faculty of Medicine. Edirne. Turkey. §Professor of Pediatrics. Department of Pediatrics. Trakya University Faculty of Medicine. Edirne. Turkey.

The study was carried out at Department of Pediatrics, Trakya University Faculty of Medicine. Edirne, Turkey. This study was supported by the Research Council of Trakya University (TUBAP-395) (Edirne, Turkey).

Correspondence:
Dr. M. Yazicioglu
Sukrupasa mah. Site kent, B Blok D: 13
22030 Edirne. Turkey
Tel.: + 90.284.2358312 - Fax: + 90.284.2358312
E-mail: yazicioglu@superonline.com
RESUMEN

Historial: La contribución al asma infantil a causa de la exposición a hongos de interior no está totalmente clara.

Objetivo: Intentamos investigar la flora de hongos que se encuentra en el aire dentro de los hogares de los niños asmáticos y los niños controlados, así como determinar la influencia de las características de la casa respecto a los hongos de interior.

Métodos: Cuarenta y siete niños asmáticos y veintiséis niños controlados no alérgicos. La reacción alérgica se determinó mediante pruebas de alergia. Se llevó a cabo una evaluación exhaustiva utilizando un cuestionario y encuestas de inspección. Las visitas domiciliarias fueron realizadas entre octubre de 2000 y febrero de 2001. Las muestras de las esporas de hongos aerotransportadas fueron recogidas en cuatro habitaciones por el método de la “placa de Petri abierta”. Se midió la temperatura y la humedad interior.

Resultados: El número total de hongos de interior en las salas de estar y en los dormitorios era notablemente más elevado en el grupo de asmáticos que en el otro grupo (p = 0,012 y p = 0,003, respectivamente). El género aislado más común fue el Cladosporium. Doce de los pacientes asmáticos (25,53 %) resultaron ser sensibles a los hongos. La regresión logística puso en evidencia que las manchas de hongos aerotransportadas fueron recogidas en cuatro habitaciones por el método de la “placa de Petri abierta”. Se midió la temperatura y la humedad interior.

Conclusión: El número de hongos de la colonia era más elevado en los hogares de niños asmáticos que en los controlados. Por lo tanto, la exposición a hongos de interior puede contribuir al desarrollo del asma infantil. Los cuartos de baño eran la fuente principal de propagación de hongos. Las casas viejas eran más propensas al crecimiento del hongos.


INTRODUCTION

Environmental factors such as indoor and outdoor air pollution, tobacco smoke and early exposure to inhalant allergens play roles in the development of asthma in genetically predisposed children1,2. Strong correlations have been found between asthma and early exposure to house dust mite3,4, pet5,6, and cockroach allergens7,8. However, the contribution of fungi to respiratory allergic diseases is not fully evaluated9, though they are the most diverse particles in the air we breathe.

In a previous questionnaire based study of 682 school children, dampness in the home and stuffed toys in the child’s bedroom were significant risk factors for asthma by multiple regression analyses (O.R = 2.61; 95 % CI 1.13 to 6.81 and O.R = 2.18; 95 % CI 1.27 to 3.74, respectively)9. Based on these data, we examined the hypothesis that indoor humidity and mold growth in house are risk factors for asthma. Limitations for our initial epidemiological evaluations were allergy skin prick tests (SPT) were not performed, true humidity in homes was not assessed, and indoor pets, toys, etc. were not verified. These factors integrated into new hypothesis.

MATERIAL AND METHODS

Patients

All files of asthmatic children, aged 4-17 years, who had visited the Allergy Clinic of Trakya University Medical School Department of Pediatrics (Edirne, Turkey) at least once during the previous year, were retrospectively analyzed. The patients were part of a large study to examine allergen sensitivities in Edirne and surrounding 100 km area of European Turkey outside Istanbul (n = 400). Among 104 children skin prick tested residing in the Edirne city center, subset of atopic subjects (n = 50) who agreed to participate in this mold evaluation study were included into the study. The families were living in the same address at least once during the previous year. They were excluded, leaving n = 23 homes for control evaluaciones.

Asthma was diagnosed in children by history, symptoms, physical findings, and the presence of reversible airflow obstruction as demonstrated by an improvement in FEV1 of greater than 12 % after bron-
Skin prick tests

Skin tests were performed with Dermatophagoides pteronyssinus and D farinae, fungi (Alternaria tenuis, Aspergillus mix, Candida albicans, Mucor racemosus, mold mix I-Alternaria tenuis, Aspergillus mix, Cladosporium, Penicillium mix, and mold mix II-Rhizopus, Mucor, Cladosporium, Penicillium, and animal dander and feathers (cat, dog, sheep wool and feathers mix), and common grasses, weeds, and trees, histamine, and saline (Stallergenes, France). The reactions were recorded after 20 minutes and a wheal diameter 3 mm larger than the wheal diameter of the negative control was considered as positive reaction. All the children tested had positive control wheals (St 11 mm, P 7 mm). None of the cases was under treatment that might modify skin tests, and none of the patients had received specific immunotherapy. Atopy was defined as having a positive SPT to at least one of the allergens in the panel. Fungal sensitization was defined as having a positive SPT to at least one of the fungi in the panel.

Questionnaire

An interviewer-administered questionnaire was used to characterize any environmental and behavioral variables that affect the presence of indoor fungal propagules. The questionnaire was administered to the parents while the family visited the hospital for the children’s skin-prick testing. The questionnaire included structural items such as age of the house, years of occupation, number of occupants, and number of rooms in the house. The instrument also surveyed environmental controls including method of heating, presence of air conditioning, humidifiers, fans, air circulation, and sun exposure. Construction materials like insulation, flooring, and wallpaper were also noted. The presence of indoor pets, cockroach infestations, carpeting, and plants was also noted. Household vapor production was assessed by noting the presence of aquariums, weekly plant watering and floor mopping frequencies, daily cooking hours, and time spent on laundry and bathing per week. The presence of items in the child’s room that tend to gather dust was also ascertained. Residents were asked to include any observation of moisture problems, damp patches, mold or mildew growth, musty odors, and flooding or water damage within the past year. Finally, the family’s socioeconomic status was determined.

Home visits and sample collection

To verify reports of dampness or mold, all homes were visited by researchers, blinded to the children’s case-control status. The surveyors looked for evidence of (a) water damage and flooding intrusion, (b) visible mold growth in any room, (c) musty odors, and (d) visible dust.

Airborne fungal spores were collected from bedrooms, living rooms, kitchens and bathrooms by exposing a petri dish with Rose-Bengal streptomycin agar. Samples were taken between 01.30 P.M and 02.30 P.M. Petri plates were put on a table 80-90 cm in height during sampling for ten minutes. The residents were asked to keep the windows and doors closed at least one hour before the measurements were taken. After sampling, the petri plates were incubated at 25 °C-27 °C for 6-7 days and the number of colony-forming units (CFU) was counted. At each sampling date, relative humidity and temperature were recorded by using a thermo hygrometer device (TFA-Dostmann GmbH, Germany) in children’s bedrooms. Three other measurements were made by the parents after training them. Home visits were made between October 2000 and February 2001 (average air temperature of 8.74 °C, relative humidity 78.26 %, with mean rainfall of 37.86 mm).

Statistics

Descriptive statistics are expressed as mean ± SD, median and range. Mann-Whitney U test was used for comparison of the continuous variables. Categorical data were tested using the chi-square statistics. Correlation between fungal colony counts and indoor relative humidity and temperature was performed with Spearman’s rho. Residential characteristics related with mold growth were assessed by logistic regression analysis (forward conditional method). All statistical analyses were performed using MINITAB release 13.32 statistical software (MINITAB Inc. US).

RESULTS

There were 47 children (23 M, 15 F; median age 8) in the asthma group and 23 children (11 M, 12 F; me-
dian age 7) in the control group (table I). Of the 47 children with asthma in this study, 35 (74.47 %) had mild asthma, and 12 (25.53 %) had moderately severe asthma. The mean (± SD) duration of symptoms of asthmatic patients was 4.18 ± 3.08 years (median, 4; range, 0.42-14).

Twelve of atopic asthmatic children (25.53 %) were sensitized to at least one of the fungal allergens (or mixture of fungal allergens) that were tested. In control children SPT’s were all negative.

Fungal colony numbers in four rooms (living rooms, bedrooms, kitchens, and bathrooms) from homes of 47 asthmatic and 23 non-atopic non-asthmatic control children are described in table II. The total fungal colony counts in living rooms and bedrooms were significantly higher in asthma group than controls (p = 0.012, and p = 0.003, respectively).

All of the petri plates from homes of children with asthma exhibited fungal growth, median CFU = 13. Samples from homes of 16 control cases showed fungal growth; median CFU = 2. The number of homes with total fungal colony counts less than or equal to 2 CFU (median) was 5 in asthma group, and 12 in control group. Total indoor fungal colony counts from entire homes were significantly higher in asthma group than controls (p = 0.009 (table II).

A total of 1665 fungal colonies were isolated in 210 Petri dishes. The rate of unidentified fungal colonies was 31.89 %. The most common isolated genus was Cladosporium, followed by Rhizopus, Penicillium, Alternaria, and Aspergillus (table III).

To investigate the associations between indoor fungal growth and environmental and behavioral variables obtained from the questionnaires, and reported or observed signs of dampness, logistic regression (forward conditional method) was used. The total colony counts were grouped as ≤ 2 CFU and > 2 CFU (median as cut off value). Visible fungal patches in the bathrooms (OR = 5.75; 95 % CI 1.19 to 27.70), and the age of the house (OR = 4.24; 95 % CI 1.34 to 13.45) were associated with indoor fungal growth (table IV).

The mean indoor temperature and relative humidity was 22.29 °C and 54.07 % in homes of asthmatic

<table>
<thead>
<tr>
<th>Table I</th>
<th>Demographic characteristics of patients and controls</th>
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<tbody>
<tr>
<td>Atopic-asthma</td>
<td>Non-atopic non-asthma</td>
</tr>
<tr>
<td>(n = 47)</td>
<td>(n = 23)</td>
</tr>
<tr>
<td>Age</td>
<td>Mean ± SD</td>
</tr>
<tr>
<td></td>
<td>Median</td>
</tr>
<tr>
<td></td>
<td>Range</td>
</tr>
<tr>
<td>Sex</td>
<td>Male</td>
</tr>
<tr>
<td></td>
<td>Female</td>
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</tbody>
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<table>
<thead>
<tr>
<th>Table II</th>
<th>Comparison of total fungal colony counts (CFU) in four rooms of the homes of asthma and control groups</th>
</tr>
</thead>
<tbody>
<tr>
<td>Living room</td>
<td>Bedroom</td>
</tr>
<tr>
<td>Atopic-asthma</td>
<td>10.87 ± 37.16</td>
</tr>
<tr>
<td>Group</td>
<td>3</td>
</tr>
<tr>
<td>Median</td>
<td>0-253</td>
</tr>
<tr>
<td>Control Group</td>
<td>2.22 ± 3.74</td>
</tr>
<tr>
<td>Median</td>
<td>1</td>
</tr>
<tr>
<td>Range</td>
<td>0-17</td>
</tr>
<tr>
<td>p = 0.012</td>
<td>p = 0.003</td>
</tr>
</tbody>
</table>

<table>
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<tr>
<th>Table III</th>
<th>Mold genera and species identified in the homes of asthmatic and control children</th>
</tr>
</thead>
<tbody>
<tr>
<td>Genus/species</td>
<td>No of colonies (CFU)</td>
</tr>
<tr>
<td>Cladosporium</td>
<td>443</td>
</tr>
<tr>
<td>Rhizopus</td>
<td>234</td>
</tr>
<tr>
<td>Penicillium</td>
<td>145</td>
</tr>
<tr>
<td>Alternaria</td>
<td>32</td>
</tr>
<tr>
<td>Aspergillus</td>
<td>14</td>
</tr>
<tr>
<td>Trichoderma</td>
<td>9</td>
</tr>
<tr>
<td>Drechslera</td>
<td>5</td>
</tr>
<tr>
<td>Monodictys</td>
<td>2</td>
</tr>
</tbody>
</table>

Table III: Mold genera and species identified in the homes of asthmatic and control children.
children and 22.39 °C and 50.80 % in controls, respectively. The total CFU counts did not significantly correlate with indoor temperature or relative humidity according to Spearman’s correlation calculation (table V).

**DISCUSSION**

A number of studies from different countries have reported adverse effects of dampness and visible fungal growth in homes on respiratory health. Epidemiological studies that relied on questionnaires or questionnaires together with visual inspection surveys found a significant association between home dampness or indoor fungal growth, and wheezing, asthma or respiratory symptoms. The severity of asthma and mean number of symptoms was found to correlate with measures of total dampness and fungal growth. However, other studies reported that the presence of fungal propagules in indoor air could not be reliably predicted by home characteristics obtained by questionnaires.

In the present study, we investigated the exposure to fungi in indoor environment, in order to assess the role of fungi in asthma. We found total indoor fungal colony numbers to be higher in the homes of asthmatic children than the controls. The difference was especially significant in the children’s bedrooms and living rooms, where the children spend most of their times (p = 0.012 and p = 0.003, respectively) (table II). Our results suggest that indoor fungal exposure may contribute to childhood asthma.

In few investigations fungus exposure differences between atopic and non atopic children have been addressed. Wickman et al observed that concentrations of dust-bound fungi were lower in the homes of atopic children than controls. This observation was explained by intense house cleaning activities of the parents of atopic children. Li et al reported that the indoor fungus concentrations of asthma and control groups were higher than those in non-asthmatic atopic children in summer, but there was no difference in indoor total fungus concentrations among the groups in winter. The presence of *Cladosporium* correlated with asthma.

In our study, sampling was performed with open petri dish (OPD), a method which relies on the gravitational deposition of indoor fungal propagules. Even though with this method the quantitative estimation of propagules in a given amount of air is not possible, strong correlations were found between the results obtained with N6-Andersen and the OPD.

### Table IV

<table>
<thead>
<tr>
<th>Independent variables</th>
<th>OR</th>
<th>95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Visible fungal patches in the bathrooms</td>
<td>5.75</td>
<td>1.19-27.70</td>
</tr>
<tr>
<td>Age of the house</td>
<td>4.24</td>
<td>1.34-13.45</td>
</tr>
</tbody>
</table>

*Total colony counts are grouped as being ≤ 2 and > 2.*

### Table V

<table>
<thead>
<tr>
<th>Independent variables</th>
<th>Indoor relative humidity (%)</th>
<th>Indoor temperature (°C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total colony counts (CFU)</td>
<td>Spearman’s rho = 0.166</td>
<td>p = 0.17</td>
</tr>
<tr>
<td></td>
<td>Spearman’s rho = -0.171</td>
<td>p = 0.16</td>
</tr>
</tbody>
</table>

The most common isolated genus in our study was *Cladosporium*, followed by *Rhizopus, Penicillium, Alternaria, and Aspergillus* (table III). Similar results have been reported in other studies. To minimize indoor contamination by outdoor molds we collected samples during late fall and winter season, and while the windows were closed. Our finding of *Cladosporium* as the most prevalent genus inside homes might be related that our homes do not have sufficient insulation which allows outdoor fungi easily enter indoors.

The skin tests to fungal allergens were found to be positive in 25.53 % of our atopic asthmatic patients. In other studies the rate of skin test sensitiv-
Damp patches, mold or mildew growth, musty odors, flooding or water damage within the past year, etc. from the questionnaires were not significantly related to the measured culturable fungal propagules in indoor air in the present study. The only questionnaire found to be a predictor of indoor fungal growth was the age of the house (table IV). Older houses are more prone to water problems in combination with insufficient heating and ventilation, resulting in home dampness, as reported by Zock et al.[27] in their questionnaire-based survey. In other studies presence of dampness[21-23], musty odor[23], moldy patches[23], carpeted floors[24-25], household pets[24], limited ventilation[26,27], or infrequent vacuuming[28], were found to effect indoor fungal growth. Our results demonstrate that indoor levels of fungi did not correlate well with the questionnaire reports. Therefore visual inspections should be needed. In our study, mold spots in the bathrooms observed by the researchers was the only factor associated significantly with indoor mold growth (table IV). This suggests that bathrooms were the main source of fungal propagules. In this investigation, the indoor viable fungal propagules did not significantly correlate with indoor temperature or relative humidity (table VI). Ambient humidity may be a poor indicator of conditions in specific indoor microenvironments, where penetrating moisture or condensation on cold surfaces promotes the growth of moulds[24]. On the other hand, our measurements were made in the children’s bedrooms, however our results suggest that bathrooms were the main source of the fungal propagules.

CONCLUSIONS

Fungal colony numbers were found to be higher in the homes of asthmatic children, especially in the children’s bedrooms and living rooms, where the children spend most of their times. Our results suggest that indoor fungal exposure may contribute to childhood asthma. Indoor levels of fungi did not correlate well with the questionnaire reports. Therefore visual inspection surveys should be needed. Mold spots in the bathrooms observed by the researchers was the only factor associated significantly with indoor mold growth. This suggests that bathrooms were the main source of fungal propagules. Although, fungal allergens are thought to be less important in the etiology of asthma than house dust mites, we hypothesize that elimination of household fungi will improve asthma status. The control of indoor molds requires a concerted approach combining fungicides, measures to reduce home dampness and the removal of mold infested items as soon as possible. Moldy items, such as a basement carpet that has suffered water damage should be removed altogether. Any measures that can be taken to reduce humidity should be recommended. Air conditioners and humidifiers have also been shown to be sources of significant mold exposure. It may be concluded that measures such as frequent application of 5 % bleach into the drainages of the basements and bath-tub, and cleaning visible mold on surfaces can reduce the mold concentration of the bathroom.

REFERENCES

18. Thomsen B, Brixman J, Torun K. Adult-onset asthma is associated with self-reported mold or environmental tobacco smoke exposures in the home. Allergy 2001;56:466-8.