West Nile virus past infections in the general population of Southern Spain

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OBJECTIVE. To analyze the prevalence of past and recent infections by West Nile virus (WNV) and the risk factors associated with WNV exposure in a representative population from southern Spain.

METHODS. Sample size was established for an estimated prevalence of past WNV infections of 5 ± 2.5% in 504 subjects. A pre-stratification was performed according to age distribution and place of residence. After random telephone solicitation and acquisition of informed consent, a serum sample was collected and an epidemiologic survey performed on all participating subjects. Samples were tested with ELISA-IgG and MAC-ELISA to detect specific IgG and IgM antibodies; results were confirmed by the plaque reduction neutralization test (PRNT).

RESULTS. Prevalence of past WNV infections confirmed by PRNT in the 504 participants was 0.6%, affecting mainly older persons (mean age 65 ± 23 vs. 34 ± 22 years; \(P = 0.018\)), those living in rural areas (5.4% vs. 0% in urban areas; \(P = 0.01\)), and individuals with risk professions (prevalence 2.8% vs. 0%; \(P = 0.048\)). None of the five recent infections detected by MAC-ELISA was confirmed by PRNT.

CONCLUSIONS. These results strongly suggest past circulation and exposure of the human population to WNV in southern Spain.

Key words: Flavivirus. West Nile Virus. Seroprevalence studies. Spain.

Seroprevalencia de infecciones por el virus del Nilo Occidental en la población general del sur de España

OBJETIVO. Analizar la prevalencia de infecciones pasadas y recientes por el virus del Nilo Occidental (VNO), así como los factores de riesgo asociados con la exposición al mismo, en una muestra representativa de la población del sur de España.

MÉTODOS. El tamaño de la muestra se estableció para una seroprevalencia de infección pasada del 5 ± 2.5% en 504 sujetos, preestratificándose ésta por edad y lugar de residencia. Los voluntarios se incluyeron tras solicitud telefónica y consentimiento, y a todos ellos se les realizó una extracción de suero y una encuesta epidemiológica. Las muestras se analizaron para detectar IgG e IgM específicas, mediante enzimoinmunoenzayo (ELISA-IgG) y enzimoinmunoenzayo por captura (MAC-ELISA), confirmándose ulteriormente los resultados positivos mediante reducción-neutralización en placa (PRNT).

Finalmente se realizó un análisis de regresión logística multivariante para determinar los posibles factores asociados con la exposición al VNO.

RESULTADOS. La prevalencia de infecciones por VNO confirmadas por PRNT en los 504 sujetos incluidos fue del 0.6%, y se vieron principalmente afectadas las personas de mayor edad (edad media 65 ± 23 frente a 34 ± 22 años; \(p = 0.01\)), las que vivían en áreas rurales (prevalencia del 5.4% frente al 0% en áreas urbanas; \(p = 0.01\)), y las personas con profesiones de riesgo (prevalencia del 2.8% frente al 0%; \(p = 0.048\)). Por el contrario, ninguna de las infecciones recientes detectadas por MAC-ELISA fue confirmada ulteriormente por PRNT.

CONCLUSIONES. Estos resultados apoyan firmemente la circulación en el pasado del VNO, así como la exposición humana del mismo, en áreas del sur de España.


Introduction

The global ecology of West Nile virus (WNV) has experienced noteworthy changes in recent years. From an endemic pattern of infection in specific areas dotted with sporadic local outbreaks every 25-30 years\(^\text{1-3}\), this flavivirus has spread to non-endemic areas over the past 10 years, causing a significant burden of human and animal disease\(^\text{4-9}\). This expansion, the causes of which are not yet totally understood, has been featured by the forthcoming
of human outbreaks preceded by severe and highly mortal enzootic affecting mainly indigenous birds. An increase in WNV viremia has also been noted in these outbreaks, confirmed by human mortality ranging between 4.3% in Romania in 1996 and 16.7% in Israel in 2000. Because of these issues, WNV is now considered a paradigm of emerging pathogens.

The presence of circulating WNV strains has also increased in the Mediterranean basin in the past years, in countries geographically and climatologically closely related to Spain, such as Italy, France, Tunisia, Algeria, and Morocco. Many factors occur in our environment to enable the introduction and expansion of WNV in Spain: (a) the ubiquitous presence of wetlands, (particularly the Doñana National Park in southwestern Spain, one of the most extensive in Europe), and stable populations of various ornithofauna species; (b) the strategic geographical location for numerous migratory birds coming from endemic areas of Africa (e.g., storks and flamingos), which annually share these natural areas and wetlands with indigenous birds; and (c) recent animal and human outbreaks occurring in many bordering countries, configuring a virtual "ring" of WNV circulation around Spain.

Despite these issues, there is little data assessing the ecology and possible circulation of WNV in Sylvatic cycles occurring in our geography. With respect to humans, no cases of confirmed acute infection have been declared, and the few existing seroepidemiologic studies were performed with techniques having poor specificity (indirect hemagglutination), thus the current impact of human exposure to WNV in Spain remains uncertain. The aim of the present study was to analyze the prevalence of past and recent WNV infections and the possible risk factors associated with exposure to WNV in a representative population from southern Spain.

Inclusion criteria and sample collection
Subjects of all age groups were selected by random telephone solicitation, with the exception of the youngest age group (0-14 years). Subjects from this last group were pre-selected in our two central laboratories, which receive blood samples from the entire area (primary care, outpatient clinics, and hospital). Based on this pre-selection, we ultimately collected samples originating from primary care centers and pre-anesthetic assessment in outpatient clinics to avoid selection of hospitalized, chronically ill, or immunosuppressed children. Subsequently, legal guardians were asked by phone for informed consent to use the remaining serum for our study, and were questioned to detect possible exclusion criteria. Once informed consent had been granted, the serum samples were stored at −80°C and an epidemiologic survey was performed on all participating subjects. The data collected included gender, age, profession, place of residence, outdoor activities, travel over the past 12 months to other parts of Spain and Europe, contact with animals, and insect bites in the past month. Risk professions were defined as those involving close contact with animals (farmers, stockkeepers, veterinarians), nature and wetlands (rangers, forest workers), or mosquitoes (extermination and pest control workers). The following persons were excluded: those unable to answer the epidemiologic survey, those who had traveled to endemic areas in the previous 12 months, and persons previously vaccinated against yellow fever, hospitalized in the last 30 days, with febrile diseases in the last 30 days, or with any type of immunosuppression.

Serological techniques
Samples were processed and tested for WNV IgG by enzyme-linked immunosorbent assay (ELISA) and for WNV IgM by antibody-capture ELISA (MAC-ELISA) (Focus Technologies, Rendon, VA, USA) according to the manufacturer's guidelines. Assays were performed in duplicate. All the positive samples were retested and the final result was the mean value of all determinations. When available, IgM-positive samples were tested with a confirmatory ELISA procedure recommended by the manufacturer (background subtract procedure).

Positive or indeterminate results in both the IgG and IgM ELISA tests were assayed by a plaque reduction microneutralization test against WNV (PRNT). For this purpose, samples were tested in duplicate and assayed twice. Two-fold dilution (25 μL) of the samples ranging from 1:16 to 1:256 were placed in a 96-well tissue culture microplate (Nunc, Roskilde, Denmark) and mixed with 25 μL containing 100 tissue culture infectivity dose (100 TCID50) of a reference ELI) WNV preparation. After 1 hour of incubation at 37°C, 50 μL of a Vero R5 cell suspension containing 4x10^5 cells/well, was added to each well. Cultures were maintained for 7 days at 37°C and 5% CO2, and then fixed with a solution containing 10% formaldehyde and 1% naphthal blue-black dye. The endpoint titer was defined as the highest serum dilution that showed no cytopathic effect in 10% of cell cultures. All these procedures were performed in a level 3 biosafety facility.

For sera showing an IgG/IgM titer against WNV underwent further commercial serological analyses against other flaviviruses, including tick-borne encephalitis (TBE). The assays for dengue IgG (Panbio Diagnostics, Queensland, Australia) and for TBE IgG and IgM (Dade Behring, Deerfield, IL, USA) were based on indirect immunosorbent assay (ELISA) using recombinant antigens as antigens. The method for dengue IgM was a 2-chain capture ELISA (Panbio Diagnostics).

Definition of WNV infection
Subjects with any ELISA IgG titer were considered as having possible past WNV infection, subjects with any MAC-ELISA IgM titer were considered as having possible recent WNV infection. Those with any ELISA IgG or MAC-ELISA IgM titer, subsequently confirmed by PRNT assay, were considered as having definitive past or recent WNV infection, respectively.

Statistical analysis
Seroprevalence was determined in the overall sample, and by place of residence and age group. Continuous variables are expressed as mean±standard deviation.
TABLE 1. Epidemiological features of subjects with past West Nile virus infection in the population of southern Spain

<table>
<thead>
<tr>
<th>Subject (gender, age, place of residence, risk-profession, outdoor activities)</th>
<th>ELISA-IgG*</th>
<th>MAC-ELISA-IgM*</th>
<th>PRNT</th>
<th>Classification</th>
</tr>
</thead>
<tbody>
<tr>
<td>Male, 77 years, rural, yes, no 217VM</td>
<td>3.75</td>
<td>Negative</td>
<td>&gt;256</td>
<td>WNV past infection</td>
</tr>
<tr>
<td>Female, 38 years, rural, yes, no 254VR</td>
<td>1.79</td>
<td>Negative</td>
<td>&lt;16</td>
<td>Not confirmed</td>
</tr>
<tr>
<td>Female, 30 years, urban, yes, no 117VM</td>
<td>1.53</td>
<td>Negative</td>
<td>&lt;16</td>
<td>Not confirmed</td>
</tr>
<tr>
<td>Male, 78 years, rural, yes, no 228VR</td>
<td>3.42</td>
<td>Negative</td>
<td>&gt;256</td>
<td>WNV past infection</td>
</tr>
<tr>
<td>Male, 80 years, suburban, yes, no 7VM</td>
<td>3.33</td>
<td>Negative</td>
<td>128</td>
<td>WNV past infection</td>
</tr>
<tr>
<td>Male, 80 years, urban, yes, no 234VR</td>
<td>3.18</td>
<td>Negative</td>
<td>64</td>
<td>WNV past infection</td>
</tr>
<tr>
<td>Female, 30 years, rural, yes, no 234VR</td>
<td>1.79</td>
<td>Negative</td>
<td>&lt;16</td>
<td>Not confirmed</td>
</tr>
<tr>
<td>Male, 78 years, rural, yes, no 228VR</td>
<td>3.42</td>
<td>Negative</td>
<td>&gt;256</td>
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<tr>
<td>Male, 78 years, rural, yes, no 228VR</td>
<td>3.42</td>
<td>Negative</td>
<td>&gt;256</td>
<td>WNV past infection</td>
</tr>
</tbody>
</table>

**Cases with WNV IgG positive result**

| Male, 73 years, rural, yes, no 217VM | 3.75 | Negative | >256 | WNV past infection |
| Female, 38 years, rural, yes, no 254VR | 1.79 | Negative | <16 | Not confirmed |
| Female, 30 years, urban, yes, no 117VM | 1.53 | Negative | <16 | Not confirmed |

**Cases with WNV IgM positive result**

| Male, 38 years, urban, yes, no 199VR | Negative | <16 | Not confirmed |
| Female, 5 years, urban, no, no 215VR | Negative | <16 | Not confirmed |
| Male, 20 years, suburban, no, no 76VM | Negative | <16 | Not confirmed |
| Female, 11 years, urban, no, no 223VR | Negative | <16 | Not confirmed |
| Male, 19 years, urban, yes, no 284VR | Negative | <16 | Not confirmed |

TABLE 2. Epidemiological features of subjects with definitive past WNV infection in the population of Southern Spain, compared with those without previous exposure

<table>
<thead>
<tr>
<th>Epidemiological feature</th>
<th>Prevalence of past WNV infection</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender (male/female)</td>
<td>1.2%/0</td>
<td>NS</td>
</tr>
<tr>
<td>Risk profession (yes/no)</td>
<td>2.8%/0</td>
<td>0.04</td>
</tr>
<tr>
<td>Travel to rural areas (yes/no)</td>
<td>1.2%/0.3</td>
<td>NS</td>
</tr>
<tr>
<td>Outdoor activities (yes/no)</td>
<td>0%/0.6</td>
<td>NS</td>
</tr>
<tr>
<td>Contact with animals (yes/no)</td>
<td>0.6%/0.7</td>
<td>NS</td>
</tr>
<tr>
<td>Insect bites in past month (yes/no)</td>
<td>0.7%</td>
<td>NS</td>
</tr>
</tbody>
</table>

NS: not significant; WNV: West Nile virus.

Results

Five subjects (nearly 1% of the 504 included) presented anti-WNV IgG titers with a mean ELISA absorbance ratio of 2.72 ± 1 (table 1); none of the remaining sera had indeterminate absorbance ratios. Prevalence of definitive past WNV infection confirmed by PRNT was 0.6% (3 positive samples, with PRNT titers of 64, 128, and 256, respectively (table 1), and was significantly higher in rural areas (5.4%), than in suburban (0.6%; P = 0.09), or urban areas (0%; P = 0.01). Mean age of subjects with past WNV infection was higher than that of uninfected subjects (65 ± 23 vs. 34 ± 22 years, P = 0.01). Comparisons of demographic and epidemiological features between subjects with and without past WNV infection are shown in table 2. No differences were detected with the exception of activity in any risk profession (P = 0.048), which was associated with prior WNV exposure. Multivariate analysis detected no independent risk factors associated with past WNV infection.

Five subjects (nearly 1% of the 504 included) showed IgM titers against WNV with a mean MAC-ELISA absorbance ratio of 2.03 ± 1.15 (table 1); WNV IgG antibodies were not detected in any of these subjects, and none of the remaining sera had indeterminate absorbance ratios. None of these individuals presented specific anti-WNV neutralizing antibodies when PRNT assay was performed (table 1). Application of a background subtract procedure to four of the remaining sera also yielded negative results.

Discussion

The data from the present study show the presence of definitive past WNV infection in the general population of southern Spain, confirming exposure of the population to this pathogen. These data complete previous preliminary studies carried out in Spain with less specific techniques that reported past infection rates of 8% in the human population of the northwestern part of the country and 3.1% by ELISA and MAC-ELISA.
in small mammals collected from several parts. In addition, the findings are in accordance with the results of a similar recent study performed in northeastern areas of the country, which detected about 0.2% of confirmed WNV past infection in the general population.

Definitive past WNV infection was characterized by affecting older persons, subjects living in rural areas, and those having risk professions. This epidemiological pattern probably reflects the means of acquisition, which may have been in the former past (older people are exposed during longer periods of time), and in subjects with closer contact to the wild cycle of WNV, since rural areas are located closer to wetlands, and therefore to reservoirs and potential vectors; the same is true for persons with risk professions.

As other authors have pointed out with respect to European countries, the present study raises questions as to the absence of human outbreaks of WNV disease in Spain, despite documented circulation of the virus, population exposure to WNV, and even declaration of sporadic outbreaks affecting equine cattle in surrounding countries. This epidemiologic feature contrasts with the dynamics of WNV disease in North America, in which WNV has expanded considerably, causing more than 7,000 cases of human disease since 1999.

Several factors may be related to this difference. Natural immunization of the majority of the population due to early exposure to WNV is hardly admissible, because the prevalence rates found in the present study are far from those reported in endemic areas, which range from 8% to 15%.

Partial immunization conferring some kind of protection against WNV due to population exposure to antigenically-related flaviviruses is also unlikely, because no indigenous cases or evidence of circulation of the Japanese encephalitis virus group or dengue virus have been reported in Spain, and yellow fever was eradicated in the final years of the 20th century.

Exposure to other flaviviruses (mainly TBE virus, which is endemic in vast territories of Europe) could highlight the problem of explaining the discordance between sero-prevalences determined by ELISA (5%) and the definitive PRNT assay (0.6%) in the present study, and could also explain the recent human cases imported by Irish tourists after a visit to southern Portugal (an area close to Andalusia), whereas there is no evidence of native human cases despite detection of WNV in indigenous mosquitoes.

This explanation is not supported by our data, which showed a lack of specific antibodies against TBE virus in the sera tested. We hypothesize that the main reason may lie in climatic factors. The relatively warm, dry summer we experienced explaining the discordance between sero-prevalences determined by ELISA (5%) and the definitive PRNT assay (0.6%) in the present study, and could also explain the recent human cases imported by Irish tourists after a visit to southern Portugal (an area close to Andalusia), whereas there is no evidence of native human cases despite detection of WNV in indigenous mosquitoes. This explanation is not supported by our data, which showed a lack of specific antibodies against TBE virus in the sera tested. We hypothesize that the main reason may lie in climatic factors. The relatively warm, dry summer we experienced explaining the discordance between sero-prevalences determined by ELISA (5%) and the definitive PRNT assay (0.6%) in the present study, and could also explain the recent human cases imported by Irish tourists after a visit to southern Portugal (an area close to Andalusia), whereas there is no evidence of native human cases despite detection of WNV in indigenous mosquitoes. This explanation is not supported by our data, which showed a lack of specific antibodies against TBE virus in the sera tested. We hypothesize that the main reason may lie in climatic factors. The relatively warm, dry summer we experienced explaining the discordance between sero-prevalences determined by ELISA (5%) and the definitive PRNT assay (0.6%) in the present study, and could also explain the recent human cases imported by Irish tourists after a visit to southern Portugal (an area close to Andalusia), whereas there is no evidence of native human cases despite detection of WNV in indigenous mosquitoes.

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