Genome Announcements

Full genome sequencing of the bluetongue virus-1 isolate MKD20/08/Ind from goat in India

Karam Chand a,*, Sanchay Kumar Biswas a, Gaurav Sharma b, Arpit Saxena a, Neha Tewari a, Sonalika Mahajan b, Awadh Bihari Pandey a

a Division of Virology, Indian Veterinary Research Institute, Mukteswar Campus, Uttarakhand, India
b Project Directorate on Foot and Mouth Disease, Indian Veterinary Research Institute, Mukteswar Campus, Uttarakhand, India

Article history:
Received 6 January 2016
Accepted 20 January 2016
Available online 22 April 2016
Associate Editor: John Anthony McCulloch

Keywords:
Bluetongue virus-1
Goat
Eastern BTV toptype
Next-generation sequencing

Abstract

This communication reports full genome sequencing of the bluetongue virus-1 (BTV-1) isolate MKD20/08/Ind from goat in northern India. The total BTV-1 genome size was found to be 19,190 bp. A comparison study between the Indian isolate and other global isolates revealed that it belongs to the ‘Eastern’ BTV toptype. The full genome sequence of BTV-1 will provide vital information on its geographical origin and it will also be proved useful for comparing the Indian isolate with global isolates from other host species.

© 2016 Sociedade Brasileira de Microbiologia. Published by Elsevier Editora Ltda. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Introduction

Diseases caused by the bluetongue virus (BTV) in domestic and wild ruminants are transmitted by certain Culicoides species.1–3 BTV is the prototype species of Orbivirus in the family Reoviridae.4 The BTV genome consists of 10 linear double-stranded RNA (dsRNA) segments that code for seven structural (VP1–VP7) and four non-structural (NS1, NS2, NS3/NS3a and NS4) proteins.5,6 Currently, only 27 distinct BTV serotypes have been identified worldwide.7 BTV is endemic in India and approximately 13 different BTV serotypes (BTV-1–4, 6, 9, 10, 12, 16–18, 21 and 23) have been isolated so far.8

In this study, we report the full genome sequencing of the BTV-1 isolate MKD20/08/Ind from goat in northern India. The full genome sequence of this isolate is available in the Bluetongue Virus Repository, All Indian Network Program on Bluetongue, Indian Veterinary Research Institute, Mukteswar, India. The complete genome was sequenced by next-generation sequencing (Ion Torrent). After extracting the viral dsRNA using Tri Reagent (Sigma, St. Louis, USA), it was purified by sequential precipitation with lithium chloride. An Ion Xpress plus fragment Library Kit was used to construct the full-length cDNA library, according to manufacturer’s protocol (Rev. A, Life Technologies). After multiplexing with different bar-coded adaptors, each library was sequenced on an Ion PI...
v2 chip (Life Technologies). To establish the complete genome, overlapping sequences were assembled with the Ion Torrent server by mapping closely related genomes (BTV-1 reference genome). The mapped file was viewed with an Integrative Genomics Viewer. The average coverage depth was 127.7 and genome base coverage at 20× was 95.91%.

The segment sizes (1–10) of MKD20/08/Ind were found to be 3944, 2940, 2772, 1981, 1765, 1635, 1154, 1125, 1052 and 822 bp, respectively. These segments encode proteins with amino acid length as follows: VP1 (1302), VP2 (961), VP3 (902), VP4 (644), VP5 (526) VP6/NS4 (330/77), VP7 (349), NS1 (552), NS2 (354) and NS3/NS3a (229/216). The phylogenetic analysis revealed that MKD20/08/Ind belongs to the ‘Eastern’ BTV topotype. The full genome sequence of the BTV-1 isolate will facilitate future molecular epidemiological investigations and will also help provide vital information on its geographic origin from India and other parts of the world.

**Nucleotide sequence accession numbers**

The full genome sequence of BTV-1 isolate MKD20/08/Ind was deposited in GenBank under accession no. KU234257 to KU234266 corresponding to segment 1 through 10.

**Conflicts of interest**

The authors declare no conflicts of interest.

**Acknowledgements**

The authors would like to thank the Director of ICAR-IVRI, Project Director of ICAR-PDFMD and the AINP-BT project for providing facilities.

**REFERENCES**