Brief report

Toll-Like Receptor 2 R753Q Polymorphisms Are Associated With an Increased Risk of Infective Endocarditis

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ABSTRACT

The ability to respond to the ligands of toll-like receptors (TLR) could be affected by single nucleotide polymorphisms in TLR codifying genes. The influence of the polymorphisms TLR2 (R753Q, R677W), TLR4 (D299G, T399I) and CD14 (C-159T) was consecutively studied in 65 patients with infective endocarditis. The control group (n=66) consisted of healthy volunteers. All the polymorphisms were genotyped by means of restriction analysis after their amplification. An association between endocarditis and variants of TLR2 R753Q (P < .001) was observed, but no association with other polymorphisms was found. The TLR2 R753Q, co-dominant (odds ratio=13.33), recessive (odds ratio=9.12) and dominant (odds ratio=3.65) genotypes showed a positive association with the infective endocarditis phenotype. The polymorphism TLR2 R753Q was associated with a greater susceptibility towards the development of infective endocarditis. Further studies are required to validate these results and identify other genetic risk factors.

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INTRODUCTION

The annual incidence of endocarditis is between 15 and 60 cases per million.1,2 The prognosis is poor and has remained steady over the last 4 decades.3 Even with antibiotic therapy, the mortality rate is more than 30% in most studies, and may be as high as 70% among certain high-risk populations.4 The pathogenic mechanism by which infection of the endocardium develops is multifactorial. However, hereditary and immune factors may also contribute to its development.1,5

The toll-like receptors (TLRs) are involved in the innate immune system. To date, 11 TLRs have been identified that recognize distinct pathogen-associated molecular patterns that have been evolutionarily conserved in specific classes of microbes. These antigens include cell wall components of Gram-positive (bacterial lipoproteins and lipoteichoic acids, detected by TLR2) and Gram-negative (lipopolysaccharide [LPS], detected by TLR4) bacteria. Recognition of LPS involves an LPS receptor complex, of which CD14 and the TLR4 are important components.4

Data suggests that the ability of certain individuals to respond properly to TLR ligands may be impaired by single nucleotide polymorphisms (SNPs) within TLR genes, resulting in an altered susceptibility to infection or inflammation.5 Polymorphisms of the TLR2 R753Q gene have been linked to variations in responses to staphylococcus6 and polymorphisms of R677W are associated...
with an increased susceptibility to leprosy and tuberculosis in Asian populations. Two common mutations in the TLR4 gene have been described, TLR4 D299G and T399I. These mutations are reportedly linked to an increased risk of Gram-negative bacterial infections and septic shock. A polymorphism in the promoter region of the CD14 gene is associated with an increase in prevalence of Gram-negative bacterial infection.

We hypothesized that polymorphisms in the genes of these innate immune receptors are associated with an increase in the prevalence of infective endocarditis. Here, we examined the frequency of the TLR2 (R753Q, R677W), TLR4 (D299G, T399I), and CD14 (C-159T) polymorphisms in patients with infective endocarditis and in healthy controls.

METHODS

Patients and Controls

Sixty-five patients (57 men, 8 women; mean ± standard deviation age 63.7 ± 11.9 years, range 32-85 years) diagnosed consecutively with infective endocarditis according to the Duke criteria were studied from December 2005 to December 2008 (flow inclusion of patients: 2006, 22 patients; 2007, 17; 2008, 26). The control group (n = 66) included healthy blood donors (55 men, 11 women; age 61.7 ± 10.2 years, range 32-79 years). Patients and controls were members of a homogeneous population, all Caucasian and residents of the same region (Castilla y León, Spain). Each participant gave informed consent for the study, which was approved by the hospital’s research committee.

Genomic DNA Preparation and Quantization

Two milliliters of whole blood samples were collected in ethylenediaminetetraacetic acid-coated tubes by standard venipuncture method. Genomic DNA was extracted from whole blood samples using the Magna Pure Compact Nucleic Acid Isolation Kit I (Roche, Mannheim, Germany) following the manufacturer’s instructions.

Analysis of Polymorphisms in Genes

All polymorphisms were genotyped by restriction analysis after polymerase chain reaction (PCR) amplification. Thirty-five cycles of PCR were performed on a thermal cycler (GeneAmp9700; Perkin-Elmer Cetus, Norwalk, Connecticut, United States). Primers were synthesized (VWR International Eurolab, Barcelona, Spain). The PCR reaction consisted of 50 ng DNA, 10 pm of each primer, 10 μl PCR master mix (Promega, Madison, Wisconsin, United States) and up to 20 μl water.

TLR2 p.R677W (rs5743706) PCR primers were 5’-CAATCCCT-CCTTCAAGTTG-3’ and 5’-CAGTTCAATCTGACACCCT-3’. The cycle program consisted of 94 °C for 30 s, 58 °C for 30 s, and 72 °C for 30 s. SsI (Fermentas, Burlington, Canada).

TLR2 p.R753Q (rs5743708) PCR primers were 5’-GAAGAGAAA-CATCTAGTGCCTCCATTTTC-3’ and 5’-CTAGGACTTTATCGCAGCTCTC-3’. The cycle program consisted of 94 °C for 30 s, 49 °C for 30 s, and 72 °C for 30 s. SsI (Fermentas, Burlington, Canada) (Fig. 1).

TLR4 p.D299G (rs4986790) PCR primers were 5’-ATTTGAGACT-TACTAGTCCGGT-3’ and 5’-TTGTGAGTCTTAACTGTTGGGAAC-3’. The cycle program consisted of 94 °C for 30 s, 53 °C for 30 s, and 72 °C for 30 s. HphI (Fermentas, Burlington, Canada).

TLR4 p.T399I (rs4986791) PCR primers were 5’-TCTCAAGGT-GATTTCGGGACG-3’ and 5’-GTTCTCAAGTGATTTTGGAATG-3’.

Figure 1. Electrophoretic migration of the toll-like receptor 2p.R753Q polymorphism.

The cycle program consisted of 94 °C for 30 s, 56 °C for 30 s, and 72 °C for 30 s. Mbol (Fermentas, Burlington, Canada). CD14 c.-159C-T (rs2569190) PCR primers were 5’TACCTCCCA-CACTCTTCTT-3’ and 5’-CTCGACATTCCCTCTGTT-3’. The cycle program consisted of 94 °C for 30 s, 59 °C for 30 s, and 72 °C for 30 s. Haelll (Roche, Mannheim, Germany).

Statistical Analysis

All statistical analyses were performed using SPSS Inc. version 15.0 (SPSS Inc., Chicago, Illinois, United States). Genotype (TLR2 R677W, TLR2 R753Q, TLR4 D299G, TLR4 T399I, and 159T CD14) distribution was tested with Hardy-Weinberg equilibrium in patient and control groups. Student’s t-test was used for the comparison of numeric variables within groups and the Pearson correlation test for the correlation of numeric variables. The χ² test and Fisher’s exact test (when appropriate) were used for comparison of groups. The data were expressed as mean ± standard deviation, and statistical significance was accepted as a P-value <0.05.

Logistic regression was used to estimate the effect of TLR2 R753Q genotypes on the odds of a patient having the phenotype for infective endocarditis under three genetic models (recessive, co-dominant, and dominant), as has been done previously.

RESULTS

Endocarditis was of a community origin in 44 (67.7%), a prior cardiopathy was detected in 39 (60%), significant risk factor was identified in 31 (47.7%), and predisposing diseases as follows: diabetes mellitus 12 (18.5%), alcoholism 6 (9.2%), cancer 4 (6.1%), chronic renal failure 3 (4.6%), chronic obstructive pulmonary disease 6 (9.2%). Gram-positive bacteria were detected as a causative agent in 50 (76.9%) patients (46.1% Staphylococcus and 30.8% Streptococcus), while Gram-negative bacteria were the etiologic agent in 5 (7.7%) patients. No causative agent was isolated in 9 (13.8%) patients and 39 patients (60%) needed surgical treatment and the hospital mortality rate was 30.8% (n = 20).

Distribution of Genotypes and Allele Frequencies in Patients and Control Group

In the analysis of genes (Table 1), significant associations with infective endocarditis were seen for TLR2 R753Q, for both the genotype (P = .001) (Fig. 2) and the allele variants (P = .001). No association was found with other polymorphisms. The genotype and allele frequencies of the TLR2 R753Q polymorphism were similar to frequencies reported from other available Caucasian data.

Further analyses were undertaken, using logistic regression to evaluate the significant TLR2 R753Q variants, 3 genetic models to test for the effect of the higher-risk genotype compared with the lowest-risk genotype, and the odds of having the infective
Table 1
Distribution of Toll-Like Receptor 2, 4 and CD14 Gene Polymorphisms in Endocarditis Patients and Controls

<table>
<thead>
<tr>
<th>Gene polymorphism</th>
<th>Genotype frequencies</th>
<th>Endocarditis n = 65</th>
<th>Controls n = 66</th>
<th>Pearson $\chi^2$</th>
<th>Allele frequencies</th>
<th>Endocarditis</th>
<th>Controls</th>
<th>OR (95% CI)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>TLR2 R677W</td>
<td>CC 36 (55.4)</td>
<td>41 (62.2)</td>
<td>95 (73.1)</td>
<td>1.3393 .5119</td>
<td>C T</td>
<td>104 (78.8)</td>
<td>28 (21.2)</td>
<td>0.731 [0.413-1.291]</td>
<td>.1696 .2795</td>
</tr>
<tr>
<td></td>
<td>TC 23 (35.4)</td>
<td>22 (33.3)</td>
<td>35 (26.9)</td>
<td></td>
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</tr>
<tr>
<td></td>
<td>TT 6 (9.2)</td>
<td>3 (4.5)</td>
<td></td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>TLR2 R753Q</td>
<td>R/R 30 (46.1)</td>
<td>50 (75.8)</td>
<td>87 (66.9)</td>
<td>13.8662 .001</td>
<td>R Q</td>
<td>115 (87.1)</td>
<td>17 (12.9)</td>
<td>0.299 [0.16-0.56]</td>
<td>.0001</td>
</tr>
<tr>
<td></td>
<td>R/Q 27 (41.6)</td>
<td>15 (22.7)</td>
<td>43 (33.1)</td>
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<td></td>
<td>Q/Q 8 (12.3)</td>
<td>1 (1.5)</td>
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<tr>
<td>TLR4 D299G</td>
<td>D/D 53 (81.5)</td>
<td>60 (90.9)</td>
<td>117 (90)</td>
<td>2.6762 .2624</td>
<td>D G</td>
<td>125 (94.7)</td>
<td>7 (5.3)</td>
<td>0.504 [0.194-1.307]</td>
<td>.2049 .152</td>
</tr>
<tr>
<td></td>
<td>D/G 11 (16.9)</td>
<td>5 (7.6)</td>
<td>13 (10)</td>
<td></td>
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<td></td>
<td>G/G 1 (1.5)</td>
<td>1 (1.5)</td>
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<tr>
<td>TLR4 T399L</td>
<td>T/T 60 (92.3)</td>
<td>59 (89.4)</td>
<td>125 (96.2)</td>
<td>1.0084 .604</td>
<td>T I</td>
<td>124 (93.9)</td>
<td>8 (6.1)</td>
<td>1.613 [0.513-5.066]</td>
<td>.6811 .409</td>
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<tr>
<td></td>
<td>T/I 5 (7.7)</td>
<td>6 (9.1)</td>
<td>5 (3.8)</td>
<td></td>
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<td></td>
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<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>I/I 0 (0)</td>
<td>1 (1.5)</td>
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<tr>
<td>CD14 C-159T</td>
<td>C/C 20 (30.8)</td>
<td>21 (31.8)</td>
<td>70 (53.8)</td>
<td>0.493 .7815</td>
<td>C T</td>
<td>75 (56.8)</td>
<td>57 (43.2)</td>
<td>0.887 [0.545-1.443]</td>
<td>.2341 .628</td>
</tr>
<tr>
<td></td>
<td>C/T 30 (46.1)</td>
<td>33 (50)</td>
<td>60 (46.2)</td>
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</tr>
<tr>
<td></td>
<td>T/T 15 (23.1)</td>
<td>12 (18.2)</td>
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</tbody>
</table>

CI, confidence interval; OR, odds ratio; TLR, toll-like receptor.

Values are expressed as no. (%).

Figure 2. Prevalence of the genotypes corresponding to the endocarditis and control groups for the polymorphism toll-like receptor 2 R753Q.

endocarditis phenotype (Table 2). All three genetic models were significant, the co-dominant one carrying the highest risk. For the recessive and co-dominant models the odds ratio (OR) for genotype effects on having infective endocarditis phenotype were 9.12 and 13.33, respectively, while the OR of the dominant model was 3.65.

**DISCUSSION**

This is the first study to show that the TLR2 R753Q polymorphism is associated with a significant increased risk of infective endocarditis, and that co-dominant, recessive and dominant genotypes are associated with the infective endocarditis phenotype.

SNPs R677W and R753Q within the gene encoding TLR2 have previously been reported to carry a risk of infectious diseases.

The R753Q polymorphism was first identified by Lorenz et al. A positive association has also been reported with *Mycobacterium tuberculosis* and with acute rheumatic fever caused by Gram-positive bacteria (β-hemolytic *Streptococcus*). Previous studies are in concordance with our findings. On the other hand, in a study of 420 consecutive patients with severe S. aureus infection, Moore et al. found no association with this SNP.

Regarding the TLR2 R677W SNP, no differences were observed in the distribution of the genotypes or alleles in our study. This is consistent with data in the literature, as this polymorphism has only been identified in Asian populations with lepromatous leprosy, and not in Caucasian European populations.

In contrast, the SNPs TLR4 D299G and C-159T CD14 do have a positive association with several infectious diseases, such as septic shock due to Gram-negative bacteria. In our study, no positive correlation was found with infective endocarditis, perhaps because only 7.7% of the patients in our study had infective endocarditis due to Gram-negative bacteria.

In conclusion, we found that the polymorphism of the TLR2 R753Q leads to increased susceptibility to infective endocarditis. It is necessary to reproduce the study of population groups with different characteristics in order to prove the validity of the results and identify other genetic factors associated with an increased risk of infective endocarditis.

**Table 2**

Logistic Regression Analysis of Toll-Like Receptor 2 R753Q Genetic Variants Associated With Infective Endocarditis (All Patients, n = 131)

<table>
<thead>
<tr>
<th>Genetic variant and reference SNP</th>
<th>Recessive model</th>
<th>Co-dominant model</th>
<th>Dominant model</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>OR (95% CI)</td>
<td>P value</td>
<td>OR (95% CI)</td>
</tr>
<tr>
<td>TLR2 R753Q</td>
<td>9.12 (1.11-75.18)</td>
<td>.04 *</td>
<td>13.33 (1.59-111.92)</td>
</tr>
</tbody>
</table>

CI, confidence interval; OR, odds ratio; SNP, single nucleotide polymorphism; TLR, toll-like receptor.

OR are presented for the highest-risk genotype as compared with the lowest-risk genotype.

The number of patients (131) reflects the genotypes available. Specific genotype comparisons are listed below.

* The values are for the comparison of the QQ genotype with the combined RQ and RR genotypes.
* The values are for the comparison of the QQ genotype with RR genotype.
* The values are for the comparison of the combined RQ and QQ genotypes with the RR genotype.
of infective endocarditis, given the importance of the variability in their genetic make up.

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CONFLICTS OF INTEREST

None declared.

REFERENCES